

بِسْمِ اللّٰهِ الرَّحْمٰنِ الرَّحِیْمِ



تفریغ لاب ما یکرو

EXP .7

محاضرة:



یاسمین خلیل

الصيدلانیة:



لجان الرُفعات



اللهم علمنا ما ینفعا ، وانفعا بما علمتنا ،
وزدنا علمًا

Experiment 7

تشخيص نشاط البكتيريا ودورها قتلها عن طريق المضاد

Qualitative methods used for the evaluation of bacteriostatic activity of different antimicrobial agents

❖ Objectives:

- Evaluate the different methods of antimicrobial ^{sensitivity} susceptibility testing. ✓
- Specifically differentiate between the **qualitative methods** of antimicrobial susceptibility testing.

❖ Introduction:

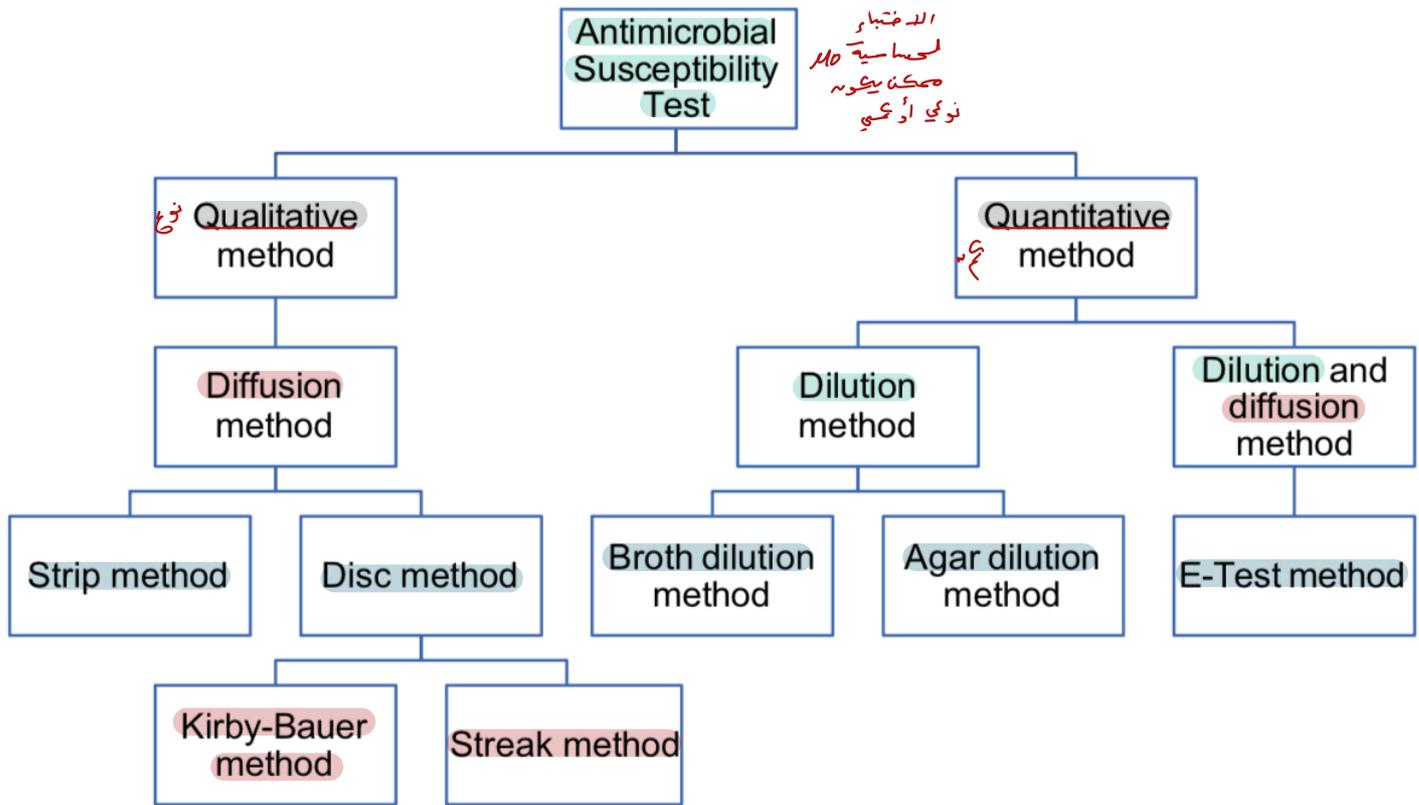
Resistance to antimicrobial agents (AMR) has resulted in morbidity and mortality from treatment failures and increased health care costs. Although defining the precise public health risk and estimating the increase in costs is not a simple undertaking, there is little doubt that emergent antibiotic resistance is a serious global problem. The results of *in-vitro* antibiotic susceptibility testing, guide clinicians in the appropriate selection of initial empiric regimens and drugs used for individual patients in specific situations (although sensitivity tests measure antimicrobial activity against bacteria under laboratory conditions it cannot be assumed that the results of *in vitro* tests will be necessarily the same *in vivo*). The selection of an antibiotic panel for susceptibility testing is based on the commonly observed susceptibility patterns, and is revised periodically.

أشياء عادية في صياغة الفقرة

التجارب في المختبرات مش شرط نتائجها تكونه بالضبط زي الجسم

اللهم ارحم ايتهم واخبره
وعافه واحق عنه وتجاوز عن
سنيته وبارك وهاغف على حسنة
وادخله وامله الجنة والمسلمين

Methods of Antimicrobial Susceptibility Testing:



هو وسط اذختر حساسیة Mo ضد المعدادات

← باستخرا diffusion

* Mueller Hinton Agar (MHA):

Mueller and Hinton developed Mueller Hinton Agar (MHA) it is commonly used for the routine susceptibility testing of microorganism by the diffusion technique.

محتویات هذا الوسط هي:

Mueller Hinton Media contains Beef Extract, Acid Hydrolysate of Casein, Starch and Agar. **Beef Extract** and **Acid Hydrolysate of Casein** provide nitrogen, vitamins, carbon, amino acids, Sulphur and other essential nutrients. **Starch** is added to absorb any toxic metabolites produced. **Starch** hydrolysis yields dextrose, which serves as a source of energy. **Agar** is the solidifying agent.

Why MHA is used for antibiotic susceptibility testing?

هاد مضافه بنقدر نن می هذا الوسط اذختر حساسیة Mo عسما نختر ماسیة مانه المعدادات

1. It is a **non-selective, non-differential medium**. This means that **almost all** organisms plated on here will grow.
كدره التشاعلی ایه ما ٧٢ المسموم يمنع تا ثر المضافه ببسکل سلبی + تنظم سیه انتشار المضافه
2. It contains starch. Starch is known to absorb toxins released from bacteria, so that **they cannot interfere with the antibiotics**. It also **mediates the rate of diffusion of the antibiotics through the agar**.
بیئته جده صیحة كانتشار المضافه + تكوين صنفقة تثقیف ببسکل اذختر صف الیئان الاذختر
3. It is a **loose agar**. This allows for better diffusion of the antibiotics than most other plates. A better diffusion leads to a truer zone of inhibition.
4. MHA shows **acceptable batch-to-batch reproducibility for susceptibility testing**.
5. MHA is low in **sulfonamide, trimethoprim, and tetracycline inhibitors** (i.e., concentration of inhibitors thymidine and thymine is low in MHA).

الذختر بی بساعه عی تقابل

الذختر نومی نتاج الاذختر هو جود صای المضافات ببسکل قابل

تستخدمها البكتيريا لكي تتاج هذا الخلية للضو

الاسيات لتكوين هذا DNA غير البكتيريا

6. Both the para-aminobenzoic acid (PABA) and thymine/thymidine content in Mueller Hinton Agar are reduced to a minimum, thus markedly reducing the inactivation of sulfonamides and trimethoprim when the media is used for testing the susceptibility of bacterial isolates to these antimicrobics.

• فالوكانه عندي كيش PABA + ثايمين في الوسط ارج تتجاوز البكتيريا تايش المضاد فارج نتون نتايج

ضا طينج حلالا مع نتونها مقاومه (PABA) مع انها في الحقيقة صباصة (موت)

* المختبر : لازم يكون PABA + ثايمين قليلا في الوسط MHA عشان دقة نتايج وصا تاثر فعالية المضاد مثل trimethoprim/sulfonamide

1] Measure length of inhibition zone

2] More than One MO

3] incubation for 24 h

← جدول هم الغرض مع الطريقة اي بعد هادي

Determination of the Susceptibility of More Than One Organisms to the Bacteriostatic Agent Provided (Strip method)

Experimental:

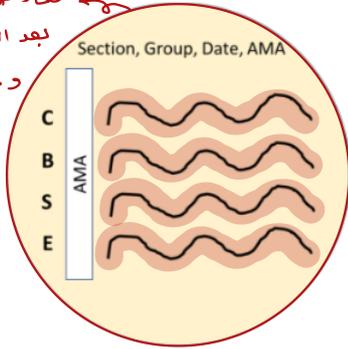
Etanol followed by heating

1. Pour an agar plate and allow to set firm
2. With a sterile forceps pick a sterile paper strip from the Petri dish and dip it into the bacteriostatic solution
3. Place the soaked strip onto the surface of nutrient agar at one side
4. Transfer a loopful from each of the test organisms to be tested and streak across the agar surface starting at the margin of the strip and at right angle to it (mark each streak on the bottom of the plate to identify)
5. Incubate at 37 °C for 24 hr.
6. Measure the length of the inhibition zones for each organism

• شريط فيه مضاد حيوي

بنسبة على أحد الجوانب في Petri dish
بمدين بنأخذ عينات MO مختلفة بدنا
نعرف مدى مقاومتها / مقاومتها لمضاد
المضاد عن طريق نقيس طول منطقة
التثبيط ومقارنتهم

عداد هو الشكل بعد الزراعة
د قبل الحضانة



AMA: Antimicrobial agent name

C: *Candida albican* Fungi

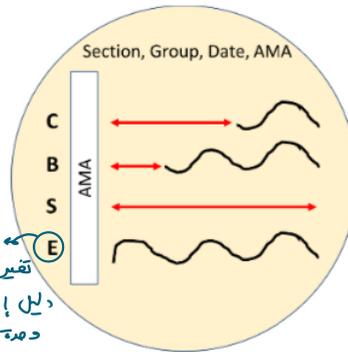
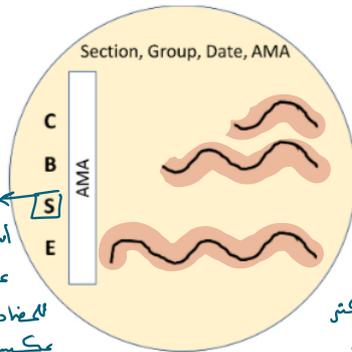
B: *Bacillus subtilis* Bacteria

S: *Staphylococcus aureus* Bacteria

E: *Escherichia coli* Bacteria

Result after incubation:

بعد 24 ساعة حضانة



كما مظهرنا امتداد MO اي اسمها S هاد دليل على شوية على انها أكثر وسعة صفة المضاد المستخدم نتم تثبيطها كلها عكسها E

تقريب في طولها دليل إنه مقاومة أكثر دعة من باقي MO

Result interpretation:

- Order according to the length of zone of inhibition:

$$S > C > B > E$$

The most sensitive

The most resistance

- So, order of organism sensitivity towards the tested antimicrobial agent is:

The most sensitive microorganism is *Staphylococcus aureus*, then *Candida albican*, then *Bacillus subtilis* and the least sensitive (most resistant) is *Escherichia coli*.

1] Measure Diameter

2] One MO

3] incubation 18-24h

➤ Determination of the susceptibility of a selected organism against different antibiotics (Kirby-Bauer method)

Kirby-Bauer (K-B) antimicrobial sensitivity test allows for the rapid determination of the efficacy of a drug (antibiotic) against certain type of bacteria by measuring the diameter of the zone of inhibition that resulted from diffusion of the drug from its disc into the surrounding medium.

K-B Test Standards:

تؤخذ العينة من مكانه صاحب فيه infection

A. Microorganisms are isolated from infected tissues. For example:

- **Septicemia**: a blood sample is obtained
- **Upper respiratory tract infections**: a throat swap or a sputum sample.

المهم هنا نحصل على نتائج صحيحة + معادلة بيدينا نوحركل اشئ في MO المزدوجة، كين؟

The obtained bacterial suspension should be **standardized** by comparing it with McFarland standard # 0.5 or # 1 (according to the type of bacteria). This would be equivalent to a bacterial concentration of 1.5×10^8 cells/mL or 3×10^8 cells/mL.

حجم هو معيار
د طرفية قياسي
كثافة البكتيريا
بمجرد (دقعة)

توحيد الوسط الزراعي كانه

B. The used medium should be **standardized**. Mueller-Hinton Agar is used. This medium with a pH of 7.2 to 7.4 contains high percent of proteins, which will facilitate diffusion from the antibiotic disc and improve bacterial growth.

تسهيل انتشار المضاد من
الاقراص وتدمع نمو البكتيريا

توحيد الاقراص الى بنانضيق مساوية MO تجا صهم

C. The antibiotic disc is made from filter paper. The concentration of certain antibiotic (antibiotic amount per disc area) should be **standardized** to ensure consistency in results. Knowing that antibiotic concentration is a factor affecting the diffusion rate from disc into the surrounding medium.

توحيد صمارة الحامضة عشرة دقة النتائج

D. The incubation temperature should be **standardized** to be 37 °C. Since temperature is also a factor affecting rate of diffusion (proportional relationship).

↑ temperature

↑ diffusion

E. The incubation time should be **standardized** to be from 18 to 24 hours (overnight incubation).

توحيد مدة الحضانة

[18-24]h

← مناسبة كلمة توصيف، بنصمكم تدرسوا أو تقرأوا

كتاب التوصيف في الإجازة

• بنضاي سطح agar مصصوة بالبكتيريا

التي هي ابيض صغار منها لا تقرأها المضاد

Experimental: و بعد نيا بنسب الأقرأها على السطح و بنسوف منقطة التقيد و بقيس قطرها

1. Prepare a bacterial lawn using MHA.
2. Mark the position for the discs with a marker pen on the bottom of the plate
3. With a sterile forceps (by ethanol followed by heating) pick a disc from the container.
4. Place the disc onto the surface of the agar at the marked position. The discs must be distributed evenly and not too close to the plate edge.
صهم نظلي صاعة حنيفة
5. Use a sterile loop to help lacing the disc on the agar.
بين الأقرأها و صا حنونة

Note:

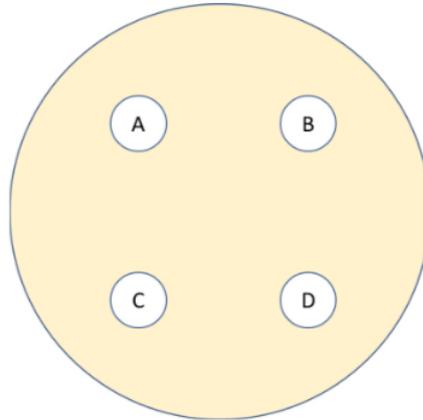
9 cm قطر Petri dish ---

إذ كانه ك الأقرأها نستعمل (Petri dish) ←

a. The minimum distance between disc centers to neighboring disc center is 24 mm, and also no closer than 10-15 mm from the edge of Petri-dish). A maximum of 6 discs may be placed on a 9-cm Petri-dish, and 12 discs on a 15-cm dish. Reduce the number of applied discs per plate if overlapping zones of inhibition are encountered.

b. A disc should not be relocated once it has come into contact with the agar surface because some of drug diffuses instantaneously. Instead, place a new disc in another location on the agar.

6. Repeat steps 3 and 4 with other antibiotics disc
7. Incubate the plate inverted at 37 °C for 24 h



الهمم يا حنونة

صونه عندكم و مط

من ردي فيه بكتيريا

و تبنا على الأقرأها

صواد و بعدين حنونة

بعد الحنونة كل ما اذنا

الحنونة الشفافة حول الأقرأها ← كل ما كانه

عدد البكتيريا المشبهة أكثر ← جلوه صا

البكتيريا صاعة لهاد المضاد (الأقرأها)

• إذا كان قطر المنطقة

صغرت ظاهرياً يسجل 0

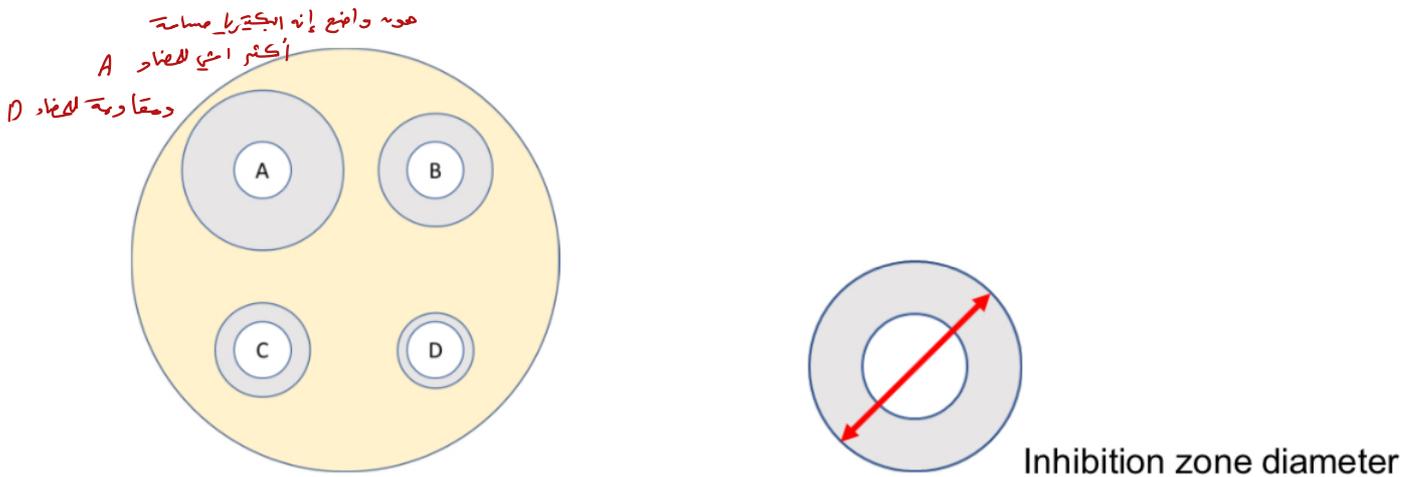
بمعنى عدم البكتيريا مقاومة

Zone of inhibition measurement and interpretation

1. After 16 to 18 hours of incubation, each plate is examined. If the plate was satisfactorily prepared, the resulting zones of inhibition will be uniformly circular and there will be a confluent lawn of growth. If individual colonies are apparent, the inoculum was too light and the test must be repeated. The diameters of the zones of complete inhibition (as judged by the unaided eye) are measured, including the diameter of the disc. Zones are measured to the nearest whole millimeter, using sliding calipers or a ruler, which is held on the back of the inverted Petri plate

Note:

If there is **NO zone at all**, report it as 0, even though the disc itself is around 7 mm.



2. The zone margin should be taken as the area showing no obvious, **visible growth that can be detected with the unaided eye**. Faint growth of tiny colonies, which can be detected only with a magnifying lens at the edge of the zone of inhibited growth, is ignored. However, discrete colonies growing within a **clear zone** of inhibition should be subculture, re-identified, and retested.
3. These zones of inhibition are then recorded for size; the diameter of the zone is proportional to the amount of anti-microbial agent, the solubility of the agent, the diffusion coefficient, and the overall effectiveness of the agent. Zones observed on the plates are then compared to standard data to determine if the isolate is truly sensitive to the antibiotic.

الهم في عناصر جمع بحسبنا إذا كانت Zone of inhibition
تأدى على ضربها للبكتيريا S. aureus حساس
Sensitive في نفس المضاد
بعدم S. aureus حساس
resist في نفس المضاد
مقاوم المرجع هو حساس

4. The sizes of the zones of inhibition are interpreted by referring to: **Zone size interpretive table for the Kirby Bauer test (CLSI table)**.

هاد جدول ظاهر ب: E. coli مع أنواع مختلفه من المضادات

Table 1: Panel of antibiotics used, their concentrations and zone diameter interpretative standards for *E. coli* (CLSI, 2007)

Group of Antimicrobial agents	Antimicrobial agents	Disk contents	Zone diameter, nearest whole (mm)			Manufacturer
			R	I	S	
Penicillin	Ampicillin	10 µg	≤ 13	14-16	≥ 17	Oxoid Ltd. Basingstoke, Hampshire, England
β-lactamase inhibitor combination	Amoxicillin-clavulanic acid	20/10 µg	≤ 13	14-17	≥ 18	
Cephems	Ceftriaxone	30 µg	≤ 13	14-20	≥ 21	
Amino glycosides	Gentamicin	10 µg	≤ 12	13-14	≥ 15	
Tetracycline	Tetracycline	30 µg	≤ 11	12-14	≥ 15	
	Doxycycline	30 µg	≤ 10	11-13	≥ 14	
Fluoroquinolones	Ciprofloxacin	5 µg	≤ 15	16-20	≥ 21	
Quinolones	Nalidixic acid	30 µg	≤ 13	14-18	≥ 19	
Folate pathway inhibitor	Trimethoprim-sulfamethoxazole	1.25/23.7 µg	≤ 10	11-15	≥ 16	
Phenicoles	Chloramphenicol	30 µg	≤ 12	13-17	≥ 18	

Classification of microorganisms according to susceptibility test results

- Susceptible (S):** ^{Sensitive} an organism is called susceptible to a drug when the infection caused by it is likely to respond to treatment with this drug, at the recommended dosage. _{يستجيب للمضاد}
- Intermediate Susceptibility (I):** covers two situations _{① مجال الاستجابة بين المقاومة والحساسية (مابين تصنيفين واهم)}

 - The organism cannot be classified as either susceptible or resistant, but is interpreted as being or intermediate (I) Susceptibility to a given drug. _{② ممكن في المضاد شغال في جسم المريض حتى بعد ما وقفه فالتصنيف I و هاد لسببين تحت}
 - The clinical interpretation of this category is that the organisms tested may be inhibited by the antimicrobial agent provided that either:

 - Higher doses of drug are given to the patient, or
 - The infection is at a body site where the drug is concentrated.
- Resistant (R):** this term implies that the organism is expected not to respond to a given drug, irrespective of the dosage and of the location of the infection.

هو اختبارنا استجابة E.coli كذا دينا

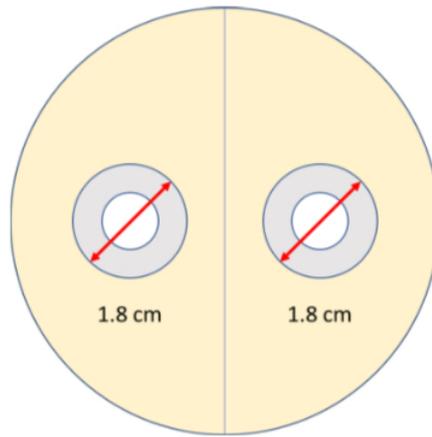
Example:

د لعلت نفس القدر !! هل نسمي مبادئ النتيجة كمنها ؟ اكبر كما بما انه مرجعي CLSI

Suppose we test the sensitivity of E-coli against ampicillin (penicillin) and ceftriaxone (cepheme).

Suppose the two zone of inhibition were 1.8 cm = 18 mm.

Compare the antimicrobial activity of ampicillin and ceftriaxone against E-coli. And classify them as a susceptible, intermediate or resistant.



بعد ما تكونوا منطقة التثبيط
وتردوا الميكون مع انهم الاقطار = 18 mm
الا انه واحدة E.coli صها كانت S
والثانية I

Table 1: Panel of antibiotics used, their concentrations and zone diameter interpretative standards for E. coli (CLSI, 2007)

Group of Antimicrobial agents	Antimicrobial agents	Disk contents	★ Zone diameter, nearest whole (mm)		
			R	I	S
Penicillin	Ampicillin	10 µg	≤ 13	14-16	≥ 17
β-lactamase inhibitor combination	Amoxicillin-clavulanic acid	20/10 µg	≤ 13	14-17	≥ 18
Cephems	Ceftriaxone	30 µg	≤ 13	14-20	≥ 21
Amino glycosides	Gentamicin	10 µg	≤ 12	13-14	≥ 15
Tetracycline	Tetracycline	30 µg	≤ 11	12-14	≥ 15
	Doxycycline	30 µg	≤ 10	11-13	≥ 14
Fluoroquinolones	Ciprofloxacin	5 µg	≤ 15	16-20	≥ 21
Quinolones	Nalidixic acid	30 µg	≤ 13	14-18	≥ 19
Folate pathway inhibitor	Trimethoprim-sulfamethoxazole	1.25/23.7 µg	≤ 10	11-15	≥ 16
Phenicoles	Chloramphenicol	30 µg	≤ 12	13-17	≥ 18

★ R: resistant, I: intermediate sensitivity, S: sensitive

هدول المشفران

اللهم انصر عبادك تحت الأتقاء
اللهم أعز الإسلام والمسلمين
اللهم ردنا بين يديك
ربّ أسألك الجنة وأعوذ بك من النار